

Olerup SSP[®] HLA - Negative Control SSP

Product number:	102.102-01 – including <i>Taq</i> polymerase
Lot number:	43K
Expiry date:	2013-April-01
Number of tests:	96
Number of wells per test:	1
Storage - pre-aliquoted primers:	dark at -20°C
- PCR Master Mix:	-20°C
- Adhesive PCR seals	RT
- Product Insert	RT

This Product Description is only valid for Lot No. 43K.

GENERAL DESCRIPTION

The *Olerup SSP[®]* HLA– Negative Control is intended to be used as a negative control in *Olerup SSP[®]* typings.

The primer set contains Negative Control primer pairs, that will amplify more than 95% of the *Olerup SSP[®]* HLA Class I, DRB, DQB1 and DPB1 amplicons as well as all the amplicons generated by the control primer pairs matching the human growth hormone gene.

The *Olerup SSP[®]* HLA – Negative Control has the sensitivity to detect approximately 50 copies of DNA template.

PRODUCT DESCRIPTION

HLA - Negative Control SSP

CONTENT

The primer set contains Negative Control primer pairs, that will amplify more than 95% of the *Olerup* SSP[®] HLA Class I, DRB, DQB1 and DPB1 amplicons as well as all the amplicons generated by the control primer pairs matching the human growth hormone gene.

PCR product sizes range from 75 to 430 base pairs.
The PCR product generated by the control primer pair is 430 base pairs.

Length of PCR product	105	200	105	80	75	80
5'-primer¹	164	340	440	45	45	43
	5'-CAC ^{3'}	5'-Agg ^{3'}	5'-TTA ^{3'}	5'-Tg g ^{3'}	5'-Tg g ^{3'}	5'-Tg g ^{3'}
3'-primer²	231	2nd I	507	59	58	57
	5'-TgC ^{3'}	5'-AAA ^{3'}	5'-TTg ^{3'}	5'-CTC ^{3'}	5'-ggC ^{3'}	5'-CTC ^{3'}
A*	+	+	+			
B*	+	+	+			
C*	+	+	+			
DRB1				+	+	
DRB3				+	+	
DRB5				+		
DQB1					+	
DPB1						+

¹The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2nd or 3rd exon, matching the specificity-determining 3'-end of the primer is given. Nucleotide and codon numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

²The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2nd or 3rd exon or the 2nd intron, matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Nucleotide and codon numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

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Lot-specific Information

www.olerup-ssp.com

The primer solution is pre-aliquoted into 0.2 ml PCR wells. Each well contains the same dried primer solution.

PCR Master Mix complete with *Taq*, *Taq* polymerase, nucleotides, buffer, glycerol and cresol red, as well as PCR lids are included in the kit including *Taq* polymerase.

PLATE LAYOUT

Each test consists of 1 PCR reaction. Each well of the 8 well PCR plates contains the same primer mix.

1	1	1	1	1	1	1	1
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The 8 well cut PCR plate is marked with ‘NC’ in silver/gray ink.

Well No. 1 is marked with the Lot No. ‘43K’.

A faint row of numbers is seen between wells 1 and 2 or wells 7 and 8 of the PCR trays. These stem from the manufacture of the trays, and should be disregarded.

The PCR plates are covered with a PCR-compatible foil.

Please note: When removing each PCR well, make sure that the remaining plates/wells stay covered. Use a scalpel or a similar instrument to carefully cut the foil between the plates/wells.

PROTOCOL

PCR AMPLIFICATION

For users of Olerup SSP® kits including Taq polymerase

Cut off one well from the 8 well PCR plate.

Add 2 µl dH₂O to the negative control well.

Add 8 µl of the PCR Master Mix complete with *Taq*-H₂O mixture to the negative control well, i.e. before the sample DNA is added to the PCR Master Mix complete with *Taq*-H₂O mixture.

Add the sample DNA to the PCR Master Mix complete with *Taq* -H₂O mixture, mix well and dispense 10 µl of the DNA-PCR Master Mix complete with *Taq*-H₂O mixture into each of the wells of the SSP typing, but not into the negative control well.

The same PCR Master Mix Complete with *Taq* and the same dH₂O that is used for the typings should be used in the negative control well. (The PCR Master Mix complete with *Taq* supplied with the Negative Control kit is intended to replace the PCR Master Mix used from the typing kits including *Taq* polymerase.)

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Lot-specific Information

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Use a 96 well thermal cycler with a heated lid. The temperature gradient across the heating block should be < 1°C.

PCR cycling parameters:

- | | | | | |
|----|------------|------|---------|-------------------------|
| 1. | 1 cycle | 94°C | 2 min | denaturation |
| 2. | 10 cycles | 94°C | 10 sec. | denaturation |
| | | 65°C | 60 sec. | annealing and extension |
| 3. | 20 cycles | 94°C | 10 sec. | denaturation |
| | | 61°C | 50 sec. | annealing |
| | | 72°C | 30 sec. | extension |
| 4. | End - hold | RT | | if less than 8 hours |
| | | 4°C | | if longer than 8 hours |

The same PCR cycling parameters are used for all the *Olerup* SSP kits.

AGAROSE GEL ELECTROPHORESIS

Prepare a 2% (w/v) agarose gel in 0.5 x TBE buffer. Dissolve the agarose by boiling in a microwave oven. Let the gel solution cool to 60°C. Stain the gel prior to casting with ethidium bromide (10 mg/ml), 5 µl per 100 ml gel solution. For maximal ease of handling use our ethidium bromide dropper bottles (Product No. 103.301-10), 1 drop of ethidium bromide solution per 50-75 ml of gel. **Note:** Ethidium bromide is a powerful carcinogen.

Load the PCR products, preferably using an 8-channel pipette. Load a DNA size marker (100 base pair ladder, Product No. 103.201-100) in one well per row.

Run the gel in 0.5 x TBE buffer, without re-circulation of the buffer, for 15-20 minutes at 8-10 V/cm.

DOCUMENTATION AND INTERPRETATION

Put the gel on a UV transilluminator and document by photography.

Record the presence and absence of PCR products.

In the negative control well no PCR product should be seen. The presence of PCR product(s) indicates contamination¹.

If contamination is detected, wipe test and testing of all reagents should be performed in order to detect the source of contamination

¹Primer oligomer artifacts may be seen. This does not represent contamination.

CERTIFICATE OF ANALYSIS

Olerup SSP[®] HLA - Negative Control SSP

Product number: 102.102-01 – including *Taq* polymerase
Lot number: 43K
Expiry date: 2013-April-01
Number of tests: 96
Number of wells per test: 1

Well specification:

Well No.	Production No.
1	2010-760-01

The negative control primer solution has been tested in a dilution series of the corresponding PCR products, 1 to 10³ down to 1 to 10⁹.

Results: The negative control primer pairs can detect contamination with the corresponding PCR products diluted 1 to 10⁷.

Date of approval: 2011-February-04

Approved by:

Quality Control, Supervisor

Lot No.: **43K**

Lot-specific Information

www.olerup-ssp.com

Declaration of Conformity

Product name: *Olerup* SSP[®] HLA - Negative Control
Product number: 102.102-01
Lot number: 43K

Intended use: Negative Control in *Olerup* SSP[®] HLA typings.

Manufacturer: *Olerup* SSP AB
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We, *Olerup* SSP AB, hereby declare that this product, to which this Declaration of Conformity relates is in conformity with the following Standard(s) and other normative document(s) ISO 9001:2008 and ISO 13485:2003, following the provisions of the 98/79/EC Directive on *in vitro* diagnostic medical devices, Annex III, as transposed into the national laws of the Member States of the European Union.

The Technical Documentation File is maintained at *Olerup* SSP AB, Hasselstigen 1, SE-133 33 Saltsjöbaden, Sweden.

The Authorized Representative located within the Community is: *Olerup* SSP AB.

Saltsjöbaden, Sweden
2011-February-04

Olle Olerup
Managing Director

Lot No.: **43K**

Lot-specific Information

www.olerup-ssp.com

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